



#### A continuous publication, open access, peer-reviewed journal

ACCESS ONLINE

### REVIEW

### Non-small-cell lung cancer: how to manage *ALK-, ROS1-* and *NTRK-*rearranged disease

#### Daniele Marinelli<sup>1,†</sup>, Marco Siringo<sup>1,†</sup>, Giulio Metro<sup>2</sup>, Biagio Ricciuti<sup>3</sup>, Alain J Gelibter<sup>1</sup>

<sup>1</sup>Division of Medical Oncology B, Policlinico Umberto I, Sapienza – Università di Roma, Rome, Italy; <sup>2</sup>Medical Oncology, Santa Maria Della Misericordia Hospital, Azienda Ospedaliera di Perugia, Perugia, Italy; <sup>3</sup>Lowe Center for Thoracic Oncology, Dana-Farber Cancer Institute, Harvard Medical School, Boston, MA, USA <sup>†</sup>These authors contributed equally to this work

#### Abstract

Oncogene addiction in non-small-cell lung cancer (NSCLC) has profound diagnostic and therapeutic implications. ALK, ROS1 and NTRK rearrangements are found in about 2-7%, 1-2% and 0.2% of unselected NSCLC samples, respectively; however, their frequency is markedly higher in younger and never-smoker patients with adenocarcinoma histology. Moreover, ALK, ROS1 and NTRK rearrangements are often mutually exclusive with other known driver alterations in NSCLC. Due to such a low frequency, diagnostic screening with accurate and inexpensive techniques such as immunohistochemistry is useful to identify positive cases; however, confirmation with fluorescent in situ hybridization or next-generation sequencing is often required due to higher specificity. In ALK-rearranged NSCLC, sequential treatment with second-generation and third-generation tyrosine kinase inhibitors leads to long-lasting disease control with most patients surviving beyond 5 years with metastatic disease. In ROS1-rearranged NSCLC, first-line treatment with crizotinib or entrectinib and subsequent treatment with

lorlatinib at disease progression leads to similar results in patients with metastatic disease. *NTRK1–3* fusions are extremely rare in unselected NSCLC. However, treatment with TRK inhibitors yields high response rates and durable disease control in most patients; diagnostic screening through multigene DNA/ RNA-based next-generation sequencing testing is therefore crucial to identify positive cases.

This article is part of the *Treatment of advanced non-small-cell lung cancer: one size does not fit all* Special Issue: https://www. drugsincontext.com/special\_issues/treatment-of-advanced-non-small-cell-lung-cancer-one-size-does-not-fit-all/

Keywords: ALK, lung adenocarcinoma, NSCLC, NTRK, ROS1, TKI.

#### Citation

Marinelli D, Siringo M, Metro G, Ricciuti B, Gelibter AJ. Nonsmall-cell lung cancer: how to manage *ALK-*, *ROS1-* and *NTRK*rearranged disease. *Drugs Context*. 2022;11:2022-3-1. https://doi.org/10.7573/dic.2022-3-1

### Introduction

Paper is divided into three sections for *ALK*, *ROS1* and *NTRK*-rearranged disease. All the available treatments are evaluated with particular attention to the specific target.

### **ALK-rearranged NSCLC**

# Clinical and biological characteristics of *ALK*-rearranged NSCLC

Anaplastic lymphoma-kinase (*ALK*) rearrangements are detectable in approximately 2–7% of patients with non-small-cell lung cancer (NSCLC) with an estimated 40,000 cases annually worldwide.<sup>1,2</sup> Patient characteristics are quite dissimilar from the overall patient population with NSCLC. They are

generally younger (median age 52 years old), are never-to-light smokers and exclusively have adenocarcinoma histology, often with signet ring or acinar histopathological features.<sup>3,4</sup> Ethnic differences amongst patients with lung cancer represent a critical issue in many aspects, including genetic characteristics, treatment response, drug toxicity and prognosis.<sup>5</sup>

ALK is a transmembrane receptor tyrosine kinase that belongs to the superfamily of insulin receptors. It activates multiple downstream pathways and may trigger neoplastic transformation. It catalyses the phosphorylation reaction of a tyrosine residue on a substrate protein that transmits ALK-mediated signals to downstream signalling pathways, even if the activation mechanism is not completely understood.<sup>6</sup> *ALK* rearrangements, which cause overexpression of a constitutively active kinase, are amongst the most common targetable alterations in NSCLC.<sup>7</sup> The most common gene alteration is an intrachromosomal inversion within the short arm of chromosome 2, joining exons 1-13 of the EML4 gene to exons 20-29 of the ALK gene; the resulting EML4-ALK fusion protein contains an N-terminal portion encoded by EML4 and a C-terminal portion (intracellular signalling portion of the receptor tyrosine kinase) encoded by ALK.<sup>8,9</sup> At least 15 EML4–ALK variants have been described in lung cancer with variants 1, 2 and 3a/b accounting for approximately 90% of them. The consequent chimeric protein activates multiple downstream known cancer signalling pathways such as PI3K-AKT, JAK-STAT and RAS-RAF-MEK-ERK. In addition, at least 20 different fusion partners have been reported, including TGF-ALK, KIF5B-ALK and STRN-ALK.<sup>10</sup> Along with other kinase fusion-positive NSCLC tumours, ALK-rearranged tumours harbour a lower tumour mutational burden than kinase fusion-negative NSCLC.<sup>11</sup>

### Molecular diagnostics of *ALK*-rearranged NSCLC

Tumour tissue sampling has traditionally been the most widely used approach to detect *ALK* translocation. Diagnosis is made using fluorescence in situ hybridization (FISH), immunohistochemistry (IHC) or next-generation sequencing (NGS) of the tumour tissue.

FISH is the diagnostic gold standard to detect rearrangements in the *ALK* locus. *EML4* and *ALK* are only separated by 12.5 megabases on chromosome 2p; therefore, FISH can be prone to false negatives when used to detect this rearrangement. Moreover, FISH is useful to determine whether there is a break in the *ALK* locus but does not distinguish between different *ALK* fusion partners.<sup>12,13</sup>

IHC is based on highly sensitive ALK antibodies to detect *ALK*positive tumours. It is an inexpensive diagnostic technique that requires less expertise and is widely available in most hospital settings, giving results faster than FISH; however, as a diagnostic tool to measure ALK protein expression on tumour cells, IHC does not identify *ALK* fusion partners.

Molecular approaches for the detection of *ALK* fusions, such as quantitative reverse transcription PCR (qRT-PCR), can facilitate diagnosis by resolving discordant or borderline cases; however, qRT-PCR is unable to detect unknown variants and fusion partners. Amplicon-based NGS is able to detect fusions with known and unknown partners or an unknown breakpoint<sup>14</sup>; NGS is also able to detect alterations in multiple driver genes at diagnosis and discriminates FISH-negative and IHC-positive cases. Moreover, tissue-based or plasma-based NGS is critical in the identification of on-target and off-target resistance mechanisms to ALK inhibitors.<sup>15,16</sup>

### Clinical activity of ALK-targeted therapies in NSCLC

Approximately 70% of patients with *ALK*-rearranged lung cancer develop intracranial metastases, with up to 30% with

intracranial disease at the time of diagnosis with significant morbidity during their disease course (Table 1).<sup>17</sup> The presence of rearrangements in the *ALK* locus renders the cancer sensitive to tyrosine kinase inhibitors (TKIs), which bind to receptor tyrosine kinases and inhibit the activation of downstream signalling pathways.<sup>18</sup> The treatment landscape for *ALK*-rearranged lung cancer has evolved rapidly over the last years (Figure 1).

#### **First-generation ALK TKIs**

#### Crizotinib

The first-in-class TKI crizotinib showed significant efficacy in the PROFILE 1001 and PROFILE 1005 (phase I and II) trials, which reported median progression-free survival (PFS) of 8–10 months amongst participants with previously treated, *ALK*-rearranged NSCLC.<sup>19,20</sup>

The first phase III trial was PROFILE 1007, which compared crizotinib with chemotherapy (pemetrexed or docetaxel) in the second-line setting in patients with locally advanced or metastatic *ALK*-rearranged NSCLC after progressing on one prior platinum-based regimen. Median PFS was longer in the crizotinib arm (7.7 *versus* 3 months, HR 0.49), and the overall response rate (ORR) was 65% *versus* 20%; overall survival (OS) was not different between treatment arms likely due to the high rates of crossover to crizotinib in patients progressing on chemotherapy.<sup>21</sup>

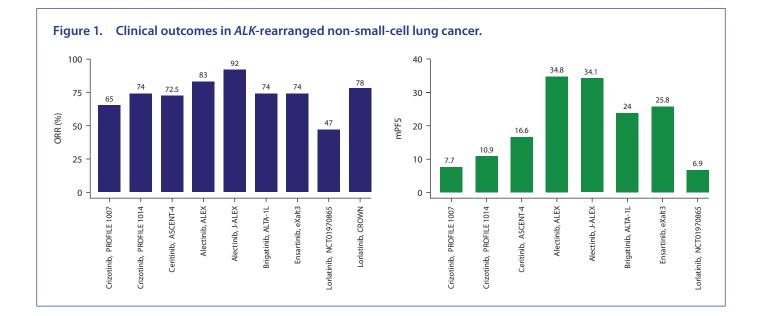
The phase III PROFILE 1014 trial compared crizotinib with standard platinum-based chemotherapy as first-line treatment for advanced *ALK*-rearranged NSCLC. Crizotinib showed longer PFS (median 10.9 *versus* 7 months in the chemotherapy arm) and higher ORR (74% *versus* 45%, respectively). Chemotherapy-related adverse events (AEs), such as fatigue, neutropenia and stomatitis, had a lower incidence in the crizotinib arm whilst low-grade vision disorders, diarrhoea and oedema were more frequent with crizotinib; however, hypertransaminasaemia was more common with crizotinib than with chemotherapy, with 14% of patients experiencing grade 3–4 AEs. A greater improvement in global quality of life (QoL) from baseline was seen amongst patients who received crizotinib than amongst those who received chemotherapy.<sup>22</sup>

Although most patients with *ALK*-rearranged NSCLC respond to crizotinib, tumours inevitably relapse, often after only 1 or 2 years of treatment because of acquired resistance. About 70% of patients with central nervous system (CNS) metastases at baseline have brain progression, whilst about 20% of patients without CNS metastases at baseline develop them. Drug failure in the CNS is linked to a pharmacokinetic issue because crizotinib is a substrate for P-glycoprotein and showed significantly lower concentrations in cerebrospinal fluid (CSF) than in plasma.<sup>23</sup>

#### Second-generation ALK TKIs

The second-generation ALK TKIs ceritinib, alectinib, brigatinib and ensartinib were developed to overcome acquired resistance to crizotinib and more efficiently penetrate the blood-brain barrier.<sup>24</sup>

ALK TKI	<b>Clinical trial</b>	Setting	Outcomes	
Crizotinib	Profile 1007	Second line after chemotherapy, crizotinib versus chemotherapy	mPFS 7.7 <i>versus</i> 3.0 months; ORR 65% <i>versus</i> 20%	
Crizotinib	Profile 1014	First line, crizotinib <i>versus</i> chemotherapy	mPFS 10.9 versus 7.0 months; ORR 74% versus 45%; mOS not reached versus 47.5 months	
Ceritinib	Ascent 4	First line, ceritinib <i>versus</i> chemotherapy	mPFS 16.6 versus 8.1 months; ORR 72.5% versus 26.7%	
Alectinib	ALEX	First line, alectinib <i>versus</i> crizotinib	mPFS 34.8 <i>versus</i> 10.9; mOS not reached <i>versus</i> 57.4 months; ORR 83% <i>versus</i> 75%	
Alectinib	J-ALEX	First line, alectinib <i>versus</i> crizotinib	mPFS 34.1 <i>versus</i> 10.2; mOS not reached <i>versus</i> 43.7 months; ORR 92% <i>versus</i> 79%	
Brigatinib	ALTA-1L	First line, brigatinib <i>versus</i> crizotinib	us mPFS 24.0 versus 11.1 months; mOS not reached ORR 74% versus 62%	
Ensartinib	eXalt3	First line, ensartinib <i>versus</i> crizotinib	mPFS 25.8 versus 12.7 months intracranial ORR 63.6% versus 21.1%; ORR 74% versus 67%	
Lorlatinib	NCT01970865	Second line, crizotinib and second- generation ALK TKI pre-treated	47% ORR in previously treated with 1 or more ALK TKIs; 39% ORR in previously treated with two or more ALK TKIs of 39%; mPFS 6.9 months	
Lorlatinib	CROWN	First line, lorlatinib <i>versus</i> crizotinib	mPFS not reached <i>versus</i> 9.3 months; mOS not reached	



#### Table 1. Clinical activity of ALK TKIs.

#### Ceritinib

Ceritinib is an ATP-competitive, highly selective ALK inhibitor and a potent inhibitor of IGFR1, ROS1 and insulin receptor but is not an efficient inhibitor of c-MET.<sup>25</sup> It is 20 times more potent than crizotinib against ALK and is effective against the L1196M, G1269A, C1156Y, I1171T and S1206Y *ALK*-resistance mutations.<sup>26</sup>

In the ASCEND-4 trial, ceritinib was compared to chemotherapy in patients with untreated, advanced *ALK*-

rearranged NSCLC showing 72.5% ORR compared to 26.7% with chemotherapy, a median PFS of 16 months compared to 8 months with chemotherapy, and 73% intracranial response rate in patients with brain metastases at baseline. However, due to an unfavourable toxicity profile because of gastrointestinal side-effects and liver toxicity, ceritinib is not listed as a preferred option in the first-line setting.<sup>27,28</sup>

#### Alectinib

Differently from crizotinib, alectinib does not inhibit MET and ROS1 but inhibits RET with a similar potency to ALK, which is five times higher than crizotinib.<sup>29</sup> It targets several *ALK* mutations that confer resistance to crizotinib (L1196, the most common mutation in crizotinib-resistant specimens, C1156Y, F1174L, R1275Q and G1269A) and has improved penetration in the CNS because it is not a substrate of P-glycoprotein. Alectinib is a preferred choice in the first-line setting in *ALK*rearranged NSCLC, but it is active both in crizotinib-naive and crizotinib-resistant *ALK*-rearranged tumours.

Alectinib was compared to crizotinib in Japanese patients in the ALEX trial (J-ALEX) showing a 92% ORR versus 79% with crizotinib and improved PFS (not reached versus 10.2 months).<sup>30</sup> These results were confirmed in the Asian ALESIA trial and the global phase III ALEX trial, which compared alectinib with crizotinib, confirming a median PFS of 34.8 versus 10.9 months and an ORR of 83% versus 75%, respectively; the intracranial response rate was 81% with alectinib, with 38% of patients showing intracranial complete responses, and 50% with crizotinib.<sup>31,32</sup> In the final results of the ALEX trial, the OS benefit of alectinib was evident across all patient subgroups with a 5-year OS rate of 62.5% versus 45.5%.<sup>33</sup> Patients treated with alectinib had a lower incidence of CNS progression both in patients with and without baseline brain metastases; moreover, the intracranial response rate in patients with prior radiotherapy was 85.7% with alectinib and 71.4% with crizotinib, whilst it was 78.6% and 40% in patients without prior radiotherapy, respectively.<sup>34</sup> Patients treated with alectinib had low rates of grade 3–4 hypertransaminasaemia and an overall lower incidence of nausea, vomiting and diarrhoea when compared to patients treated with crizotinib.<sup>32</sup> Furthermore, patients treated with alectinib showed improvements in lung cancer symptoms for longer than patients treated with crizotinib, with longer duration of clinically meaningful improvements in health-related QoL and better patientreported tolerability.35

#### Brigatinib

Brigatinib is a TKI with activity against ALK, ROS1, IGF1R and FLT3 as well as EGFR deletions and point mutations; it showed activity against multiple *ALK*-resistance mutations and a 12-fold higher potency than crizotinib against ALK.<sup>36</sup> Brigatinib showed efficacy both in treatment-naive and in crizotinib-resistant ALK-rearranged tumours.<sup>37</sup>

In patients with crizotinib-resistant, *ALK*-rearranged NSCLC, ORR was 62%, median PFS was 14.5 months and duration of response was 11.2 months.<sup>38</sup>

First-line brigatinib was compared with crizotinib in untreated, *ALK*-rearranged NSCLC in the phase III ALTA-1L trial; grade 3–4 increased blood creatine kinase levels, hypertension and increased lipase levels were more common with brigatinib than with crizotinib.<sup>39</sup> In the final results of ALTA-1L, brigatinib showed median PFS of 24 *versus* 11 months for crizotinib,

median intracranial duration of response in patients with measurable brain metastases at baseline of 27.9 *versus* 9.2 months with crizotinib, and the 3-year intracranial PFS rate was 31% with brigatinib and 9% with crizotinib (HR 0.29).<sup>40</sup> Healthrelated QoL and multiple functional and symptom scales were improved in patients treated with brigatinib in the ALTA-1L trial when compared to patients treated with crizotinib.<sup>41</sup>

#### Ensartinib

Ensartinib is a second-generation small-molecule TKI that selectively inhibits ALK with a potency more than 10 times greater than crizotinib. It also potently inhibits most common crizotinib-resistance mutations, including F1174, C1156Y, G1269A, L1196M, S1206R and T1151.<sup>42</sup>

Objective response with ensartinib was achieved in 52% of patients and median PFS was 9.6 months in patients with crizotinib-resistant, *ALK*-rearranged NSCLC.<sup>43</sup>

The eXalt3 randomized phase III trial compared ensartinib with crizotinib amongst patients with previously untreated *ALK*-rearranged NSCLC. In the intention to treat population, the median PFS was significantly longer with ensartinib than with crizotinib (25.8 *versus* 12.7 months respectively, HR 0.51) and the intracranial response rate was 63.6% *versus* 21.1%, respectively, for patients with brain metastases at baseline. About 11% of patients treated with ensartinib had grade 3 rash, which was managed by drug withholding and dose reductions.<sup>44</sup>

#### **Third-generation ALK TKIs**

#### Lorlatinib

Lorlatinib is a third-generation *ALK* inhibitor with activity against ALK and ROS1 as well as against TYK1, FER, FPS, TRK A/B/C, FAK, FAK2 and ACK; it was developed specifically to penetrate the blood–brain barrier and to overcome secondary resistance mutations emerging after treatment with firstgeneration and second-generation *ALK* inhibitors, including the G1202R mutation.

Lorlatinib has significantly improved (>50-fold) inhibitory potency and was initially tested in *ALK*-rearranged NSCLC population either with progression on crizotinib and at least one more *ALK* inhibitor or in patients with progression on either alectinib or ceritinib as first-line therapy.

Amongst 198 patients previously treated with one or more ALK inhibitors, ORR with lorlatinib was 47%; amongst patients who had failed two or more ALK TKIs, ORR was 39% and median PFS was 6.9 months. Furthermore, lorlatinib showed improved efficacy in patients with *ALK*-resistance mutations, with the most common being the *ALK* G1202R mutation; those findings suggested higher lorlatinib activity in *ALK*-rearranged tumours that have acquired on-target resistance mechanisms.<sup>45</sup>

Recently, the phase III randomized CROWN trial compared lorlatinib with crizotinib in 296 patients with untreated, advanced and *ALK*-rearranged NSCLC. The results showed that median PFS was not reached with lorlatinib *versus*  9.3 months with crizotinib (HR 0.28); at 12 months, the percentage of patients alive and progression-free at 12 months was 78% with lorlatinib and 39% with crizotinib; ORR was 76% versus 58%, respectively. Intracranial response rate in patients with measurable brain metastases at baseline was 82% with lorlatinib and 23% with crizotinib; the rate of intracranial PFS at 12 months was 33.2% and 2.8%, respectively. Lorlatinib showed the highest rates of intracranial efficacy, including an intracranial complete response rate of 61%. For patients without baseline brain metastasis, the intracranial control rate with lorlatinib at 12 months was 97%. Lorlatinib showed significant intracranial activity and clinically meaningful benefit also in previously irradiated brain lesions that were in progression at baseline. Patients treated with lorlatinib had a greater improvement from baseline QoL than patients treated with crizotinib. Patients treated with lorlatinib had a higher incidence of grade 3-4 hypertriglyceridaemia, hypercholesterolaemia, and increased weight and hypertension; moreover, mood effects, such as anxiety, depression and others, were highlighted in 9% of patients treated with lorlatinib and were most common in the first 2 months of lorlatinib administration.<sup>46</sup>

# Mechanisms of resistance to ALK inhibitors

As with most targeted therapies, resistance mechanisms to *ALK* inhibitors are broadly divided into on-target and off-target mechanisms.

As already discussed, whilst CNS progression on crizotinib may often be due to its lower concentrations in the CSF, on-target resistance mutations or ALK gene amplification typically arise in about one-third of crizotinib-resistant samples and in about half of alectinib/ceritinib-resistant specimens.<sup>47</sup> On-target mutations highlight persistent ALK oncogenic activation evolving through different resistance mutations in time and space under selective pressure imposed by different ALK TKIs.<sup>48</sup> Accordingly, lorlatinib efficacy is improved in patients with ontarget mutations arising after treatment with first-generation or second-generation ALK TKIs; in more detail, lorlatinib retains activity against the G1202R mutation - the most common resistance mutations arising after treatment with secondgeneration ALK TKIs.<sup>49</sup> However, sequential treatment with ALK inhibitors fosters the development of ALK compound mutations, leading to resistance to all available inhibitors.<sup>50</sup> Compound mutations have the utmost frequency in extensively pre-treated, lorlatinib-resistant tumour specimens; nonetheless, off-target oncogenic drift due to the acquisition of bypass mechanisms of resistance can, at any time, override ALK-centred oncogene addiction, leading to multiple hard-totreat resistance patterns.<sup>51</sup> Fourth-generation ALK inhibitors are being developed to retain activity against compound ALKresistance mutations.<sup>52</sup> Different fusion variants may also shape sensitivity to ALK TKIs and the emergence of ALK-resistance mutations.53-56

# Suggested approach in *ALK*-rearranged NSCLC

First-line treatment with second-generation ALK TKIs is critical due to the highly relevant impact on survival and patientrelated outcomes shown in multiple comparisons with crizotinib in phase III clinical trials. In case of asymptomatic CNS metastases, local therapy can often be deferred due to high intracranial response rates. However, local ablative therapies are a major treatment option both in cases of oligoprogressive disease in the brain and in cases of non-CNS oligoprogression, with the aim of maintaining the benefit from each *ALK*-directed line of treatment and to prolong time to chemotherapy.

Whilst lorlatinib showed major efficacy in the first-line setting, in the absence of head-to-head comparisons with second-generation ALK TKIs and whilst waiting for mature survival data, its most appropriate use may be after resistance to second-generation TKIs. However, results from the phase III CROWN study showed the highest efficacy amongst all *ALK*-directed TKIs; therefore, the optimal choice for first-line therapy in *ALK*-rearranged lung cancer remains unclear. As of May 2022, the NCCN NSCLC Panel lists alectinib, brigatinib and lorlatinib as preferred options for patients with *ALK*-rearranged metastatic NSCLC.<sup>57</sup> Drug availability and drug pricing are an issue both in western and developing countries and can influence the choice of treatment, whilst long-term management of AEs is critical.

*ALK* tumours displayed poor sensitivity to single-agent immune-checkpoint inhibitors likely due to an unfavourable microenvironment.<sup>58</sup> However, sensitivity to platinum/ pemetrexed combination chemotherapy is retained both in tumours that are TKI naive and in those pre-treated with TKIs, and chemotherapy is a viable alternative at progression from all available ALK TKIs.<sup>59,60</sup> Because *ALK*-rearranged tumours were excluded from most chemoimmunotherapy trials, chemotherapy/immunotherapy combinations are not indicated at the progression from ALK-directed therapies.<sup>61</sup>

Liquid biopsy was shown to be able to track the evolution of resistance to TKIs during therapy; blood-based monitoring of resistance mechanisms can provide critical information on treatment sequencing. However, broad standardization of techniques to monitor resistance is lacking, and such efforts should be limited to centres with significant expertise.

### **ROS1-rearranged NSCLC**

The proto-oncogene *ROS1* encodes a receptor tyrosine kinase with an unclear role in human physiology whose kinase domain is highly homologous with *ALK*; approximately 1–2% of NSCLC harbour *ROS1* rearrangements.<sup>62,63</sup> In the last 10 years, multiple TKIs have shown efficacy in *ROS1*-rearranged NSCLC, significantly reshaping the therapeutic landscape for patients harbouring this aberration.

# Clinical and biological characteristics of *ROS1*-rearranged NSCLC

ROS1 rearrangements in NSCLC occur predominantly in younger patients with adenocarcinoma histology and light or no smoking history; large cell and squamous histology are uncommon, and median age at diagnosis is 50 years.<sup>64</sup> The spectrum of incidence is highly overlapping with ALKrearranged NSCLC; in particular, ROS1 fusions were described in 2.2% of never smokers, whilst ALK rearrangements were described in 5.6% of patients in the same cohort.<sup>65</sup> About a third of treatment-naive patients with metastatic disease have brain metastases, and progressive disease in the CNS is found in up to 50% of patients pre-treated with TKIs.<sup>66</sup> CD74-ROS1 fusions displayed a higher frequency of CNS metastases when compared to non-CD74 fusion partners; however, it is unclear whether fusion type affects CNS spread.<sup>67</sup> Moreover, venous thromboembolic events are described in about 40% of patients with ROS1-rearranged NSCLC and are more frequent than in unselected patients with NSCLC;<sup>68,69</sup> thus, specific attention to signs and symptoms of deep vein thrombosis and pulmonary embolism is needed in this subpopulation.

Heterogeneity in partner genes is described in solid tumours harbouring *ROS1* rearrangements; however, the most common fusions in NSCLC are *CD74–ROS1* (found in about 44% of patients), *EZR–ROS1* (16%), *SDC4–ROS1* (14%) and *SLC34A2– ROS1* (10%). The pattern of structural rearrangement involves loss of the extracellular *ROS1* domain and fusion of the kinase domain with the N-terminal portion of a partner gene.<sup>70</sup> *ROS1*-rearranged tumours harbour a low tumour mutational burden;<sup>71</sup> the co-occurrence of *ROS1* rearrangements with *EGFR* or *KRAS* mutations and *ALK* rearrangements in the same tumour is rare.<sup>72</sup> Whilst *ROS1*-rearranged tumours may seldom show high PD-L1 expression, the efficacy of immune-checkpoint inhibitor monotherapy is likely to be modest.<sup>73</sup>

### Molecular diagnostics of *ROS1*-rearranged NSCLC

Due to the rarity of *ROS1* rearrangements and the limitations associated with any diagnostic test, false-positive and false-negative results may occur.<sup>74</sup>

Break-apart FISH is regarded as a diagnostic gold standard: split probes binding to the 5' and 3' ends of *ROS1* or isolated 3' signals in more than 15% of tumour cells on a minimum of 50 cells identify positive cases.<sup>74</sup> RT-PCR detects known fusion patterns through specific primers;<sup>75</sup> both FISH and RT-PCR were used to identify eligible patients in seminal trials.<sup>70</sup> IHC is a useful screening tool for *ROS1* rearrangements due to its high sensitivity and specificity; however, confirmation by FISH or RT-PCR/NGS is required after IHC positivity, whilst IHC-negative cases can be interpreted as negative for *ROS1* rearrangements.<sup>76–83</sup> DNA-based NGS is able to identify cases with negative results on non-NGS testing;<sup>84</sup> RNA-based tests may further increase sensitivity due to the lack of coverage of introns inferred to be the site of the genomic breakpoints by DNA-based NGS and because high expression of the fusion mRNA can mitigate false-negative results of DNA-based NGS due to low tumour purity.<sup>85</sup>

# Clinical activity of *ROS1*-targeted therapies in NSCLC

Crizotinib, a first-generation ALK TKI, showed efficacy against ROS1-rearranged NSCLC in the phase I PROFILE 1001 trial amongst 53 pre-treated patients, showing 72% ORR, median PFS of 19 months and a median OS of 51.4 months; median time to response was 7.9 weeks (Table 2).<sup>70,86</sup> Similar results were confirmed in a single-arm, phase II trial amongst 127 East Asian pre-treated patients, with an ORR of 71.7% and median PFS of 15.9 months<sup>87</sup> (Figure 2). Moreover, further phase II European trials (EUCROSS,<sup>88</sup> Acsé,<sup>89</sup> METROS<sup>90</sup>) confirmed an ORR of 65-70%. Median PFS was 20 and 22.8 months for EUCROSS and METROS; however, despite a similar ORR, the Acsé trial reported a median PFS of only 5.5 months in a more heavily pre-treated population with 25% of patients with ECOG PS2 amongst the ROS1-positive cohort. The intracranial efficacy of crizotinib is not well characterized in patients with ROS1-rearranged disease; intracranial ORR in the METROS trial was 33% (2 out of 6 patients). However, crizotinib CSF concentrations are low, and intracranial efficacy is inferior to second-generation and third-generation TKIs.<sup>23</sup> Accordingly, CNS is a critical and frequent site of progression in patients positive for ALK and ROS1 treated with crizotinib, even if ROS1-rearranged tumours seem to have decreased tropism for the brain when compared to ALK-rearranged tumours.<sup>91</sup>

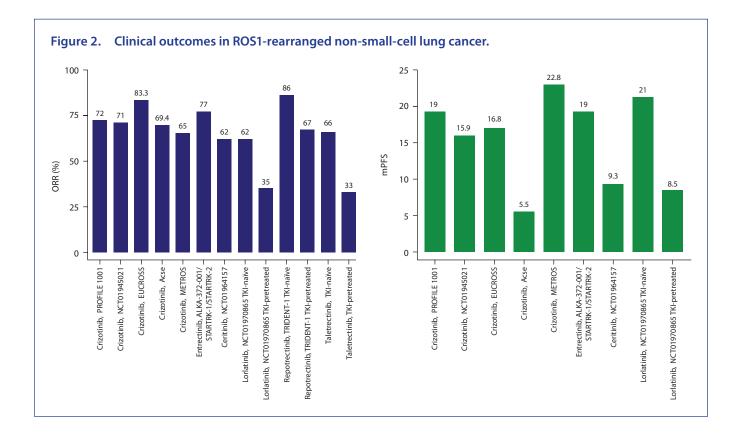
Entrectinib is a TRK A/B/C, ALK and ROS1 TKI with a 40-times greater potency than crizotinib in vitro in ROS1-rearranged cancer models.<sup>92</sup> Moreover, it was developed to efficiently cross the blood-brain barrier.93 Results from two phase I trials (ALKA-372-001, STARTRK-1)<sup>94</sup> and one phase II trial (STARTRK-2)<sup>95</sup> are available. In a ROS1 TKI-naive population, entrectinib showed a 77% ORR amongst 53 evaluable patients; amongst 17 patients with CNS metastases at baseline, intracranial ORR was 55%, median PFS was 19 months in the overall population and 13.6 months in patients with baseline CNS metastases, and in patients without baseline CNS metastases, median PFS was 26.3 months.<sup>95</sup> Thus, entrectinib compares favourably with crizotinib in a TKI-naive population due to a greater efficacy shown in patients with CNS disease; however, due to its activity against tropomyosin receptor kinase (TRK), entrectinib showed a peculiar toxicity profile causing dizziness, weight gain, paraesthesias and cognitive changes.

The second-generation *ALK* and *ROS1* inhibitor ceritinib also showed efficacy in a *ROS1*-rearranged TKI-naive population in a Korean phase II study on 32 patients;<sup>96</sup> however, due to an unfavourable toxicity profile, crizotinib and entrectinib remain the preferred options in the TKI-naive setting.

Table 2. Clinical activity of ROS1 TKIs.

ROS1 TKI	Clinical trial	Setting	Outcomes
Crizotinib	PROFILE 1001	Advanced ROS1 <sup>+</sup> NSCLC	mPFS 19 months; mOS 51.4 months 72% ORR
Crizotinib	NCT01945021	Advanced ROS1 <sup>+</sup> NSCLC	mPFS 15.9 months; 71% ORR
Crizotinib	EUCROSS	Advanced ROS1 <sup>+</sup> NSCLC	mPFS 16.8 months; 83.3% ORR
Crizotinib	Acsé	Advanced ROS1 <sup>+</sup> NSCLC	mPFS 5.5 months; mOS 17.2 months 69.4% ORR
Crizotinib	METROS	Advanced ROS1 <sup>+</sup> NSCLC	mPFS 22.8 months; mOS not reached; 65% ORR
Entrectinib	ALKA-372-001, STARTRK-1, STARTRK-2	Advanced ROS1 <sup>+</sup> NSCLC	mPFS 19 months; mOS not reached 77% ORR; 55% intracranial ORR
Ceritinib	NCT01964157	Advanced ROS1 <sup>+</sup> NSCLC	mPFS 9.3 months; mOS 24 months; ORR 62%
Lorlatinib	NCT01970865	TKI-pre-treated ROS1 <sup>+</sup> NSCLC	mPFS 21 months (TKI naive); mPFS 8.5 months (TKI pre-treated); 62% ORR (TKI naive); 35% ORR (TKI pre-treated)
Repotrectinib	TRIDENT-1	TKI-pre-treated ROS1 <sup>+</sup> NSCLC	86% ORR (TKI naive); 40–67% ORR (TKI pre-treated)
Taletrectinib	NCT02279433, NCT02675491	TKI-pre-treated ROS1 <sup>+</sup> NSCLC	66% ORR (TKI naive); 33% ORR (TKI pre-treated)

mOS, median overall survival; mPFS, median progression-free survival; NSCLC, non-small-cell lung cancer; ORR, overall response rate; TKI, tyrosine kinase inhibitor.



Lorlatinib is a third-generation ALK and ROS1 TKI with improved CNS activity due to high CSF fluid concentrations through the reduction of P-glycoprotein-mediated efflux.<sup>97,98</sup> In a phase I/ Il study, lorlatinib showed a 62% ORR amongst 21 patients with *ROS1*-rearranged disease who were TKI naive and a 35% ORR amongst 40 patients pre-treated with crizotinib.<sup>99</sup> Amongst

patients with brain metastases, lorlatinib showed intracranial ORR in 64% of 11 TKI-naive patients and in 50% of 24 patients pretreated with crizotinib. Median PFS was 21 months in TKI-naive patients and 8.5 months in patients pre-treated with crizotinib.

Repotrectinib is a next-generation TKI with efficient bloodbrain barrier penetration, developed to inhibit both wild-type and solvent-front mutations involving *ROS1, TRK A/B/C* and ALK.<sup>100,101</sup> In the phase I/II TRIDENT-1 trial, repotrectinib showed an ORR of 86% amongst 7 patients with *ROS1*-rearranged disease who were TKI naive and of 40–67% amongst patients pre-treated with TKIs.<sup>102</sup> Taletrectinib is a pan-TRK and *ROS1*-selective inhibitor active against the *ROS1* solvent-front mutation G2032R, which showed a 33% ORR and 88% disease control rate amongst six patients pre-treated with crizotinib and a 66% ORR in patients with *ROS1*-rearranged disease who were TKI naive.<sup>103,104</sup>

### Mechanisms of resistance to *ROS1* inhibitors

An on-target G2032R resistance mutation shared amongst multiple metastatic sites was first described after treatment with crizotinib in a patient with ROS1-CD74 rearranged disease in 2013.<sup>105</sup> On-target resistance mutations are the most common resistance mechanism to crizotinib, found in over 50% of patients, with ROS1 G2032R being the most frequent, causing steric interference with crizotinib and preventing effective binding. Amongst patients with CNS progression on crizotinib, it is likely that pharmacokinetic mechanisms leading to low CSF concentration may lead to resistance in the CNS rather than to on-target resistance mutations.<sup>23</sup> There were no responses to lorlatinib amongst six patients pre-treated with crizotinib with on-target resistance mutations;<sup>99</sup> ROS1 resistance mutations were also described in circulating tumour DNA samples after treatment with entrectinib.<sup>106</sup> On-target resistance mutations were also observed in 46% of cases after treatment with lorlatinib, with G2032R found in 32% of the total cases and the discovery of compound resistance mutations (G2032R-L2086F, G2032R-S1986F-L2086F, S1986F-L2000V) in the same sample.107

Off-target, *ROS1*-extrinsic resistance mechanisms were also described through downstream activating mutations on multiple kinases and epithelial-to-mesenchymal transition.<sup>108,109</sup> Moreover, spatial and temporal heterogeneity can lead to the co-occurrence of multiple resistance mechanisms in the same patient both on-target and offtarget.<sup>110</sup>

# Suggested approach in *ROS1*-rearranged NSCLC

In the first-line setting, both crizotinib and entrectinib are approved by the FDA and EMA in *ROS1*-rearranged NSCLC, and both are preferred drugs as per NCCN guidelines.<sup>57</sup> In case of metastatic CNS disease, entrectinib is preferred over crizotinib due to its higher intracranial efficacy; in case of symptomatic CNS metastases, local control with surgery or radiotherapy is needed before treatment with entrectinib. In patients with no evidence of CNS metastases, crizotinib may be preferred due to a more favourable toxicity profile; however, careful monitoring of signs and symptoms of brain metastases

is needed, along with contrast-enhanced brain computed tomography or magnetic resonance imaging. Oligoprogressive disease may be treated with local therapy. In case of systemic progression from first-line entrectinib or crizotinib, if available, lorlatinib is indicated. Enrolment in clinical trial therapy is strongly recommended for patients with *ROS1*-rearranged tumours.

### NTRK-rearranged NSCLC

Encoded by *NTRK* genes, TRKs are critical in neuronal development and functioning and act as receptors for multiple neurotrophins.<sup>111–113</sup> Chromosomal rearrangements involving *NTRK1*, *NTRK2* or *NTRK3* lead to constitutive downstream signalling and oncogenic TRK activation in a ligand-independent fashion.<sup>114,115</sup> Whilst oncogenic *NTRK* rearrangements are pathognomonic amongst specific rare cancers, such as secretory cancers of the salivary gland, their frequency amongst the most common solid tumours is extremely low.<sup>116</sup>

# Clinical and biological characteristics of *NTRK*-rearranged NSCLC

NTRK1-3 rearrangements arise in 0.17-0.23% of unselected NSCLC, more commonly in non-smokers with adenocarcinoma histology and young age (median age 48 years); however, NTRK rearrangements are also identified in older patients or in patients with squamous cell or neuroendocrine histology.<sup>117-119</sup> Similarly to ALK-rearranged and ROS1-rearranged tumours, NTRK-rearranged tumours exhibit a low tumour mutational burden.<sup>11</sup> NTRK1 fusions were described in 3.3% of NSCLC samples negative for oncogenic alterations.<sup>120</sup> Rearrangements occur through fusion of the 3' NTRK1, NTRK2 or NTRK3 sequence with the 5' sequence of a partner gene; partner genes are frequently characterized by oligomerization domains, which contribute to the oncogenic potential of the chimeric protein.<sup>121–123</sup> The first to be described and most common fusion partner in adult NTRK-fusion-positive tumours is ETV6, followed by many others (TPM3, TPR, SQSTM1, IRF2BP2).124

# Molecular diagnostics of *NTRK*-rearranged NSCLC

Given the rarity of *NTRK1*, *NTRK2* and *NTRK3* rearrangements in NSCLC, broad molecular profiling through DNA-based or RNAbased NGS testing for multiple alterations is critical. Known fusions can be efficiently detected through amplicon-based DNA sequencing, whilst hybrid capture library preparation is able to detect both known and novel fusion partners; however, rearrangements involving intronic regions can lower DNA-based NGS sensitivity. DNA-based sequencing can detect chromosomal rearrangements, which may or may not lead to functional fusion transcripts, whilst RNA-based NGS is a critical tool for diagnosis of de novo, transcribed gene fusions.<sup>125</sup> Anchored multiplex PCR, amplicon-based multiplex PCR and hybrid capture-based RNA NGS provide high sensitivity and concordance and can detect gene rearrangements in samples that appear negative for driver mutations after DNA-based NGS.<sup>126</sup> Nevertheless, the labile nature of RNA in formalin-fixed paraffin-embedded archival samples may lead to false-negative results. A parallel or sequential DNA-based and RNA-based NGS approach maximizes diagnostic sensitivity and appropriate evaluation of driver comutations.<sup>127</sup> IHC screening is an inexpensive diagnostic tool; however, IHC staining patterns are not specific for *NTRK* rearrangements and can detect wild-type TRK expression. In case of positive results, confirmation with FISH or NGS is required.<sup>128,129</sup>

# Clinical activity of *NTRK*-targeted therapies in NSCLC

Larotrectinib is a first-in-class, highly selective *TRK A/B/C* inhibitor showing 75% ORR in *TRK*-fusion-positive cancers in adults and children.<sup>130</sup> Amongst 20 heavily pre-treated patients with *TRK*-positive NSCLC with a median age at diagnosis of 48 years, larotrectinib showed a 73% ORR amongst 15 evaluable patients and 63% intracranial ORR amongst 8 evaluable patients. Median OS was 40.7 months.<sup>131</sup>

As previously discussed, entrectinib is active both against *ALK/ ROS1* and *NTRK*-rearranged tumours. Amongst 54 adults with advanced *NTRK*-positive solid tumours entrectinib showed 57% ORR.<sup>132</sup> Amongst 13 patients with *NTRK*-positive NSCLC, entrectinib showed a 69% ORR with a median PFS and OS of 14 months,<sup>133</sup> amongst 8 patients with baseline CNS metastases, entrectinib showed a 63% intracranial ORR.<sup>134</sup>

As the TRK pathway is involved in appetite, balance and pain sensitivity, *TRK* inhibitors frequently lead to on-target AEs such as dizziness, weight gain, withdrawal pain and paraesthesias. Weight gain and pain upon *TRK* inhibitor withdrawal were associated with longer treatment exposure, whilst dizziness showed a median time to onset of 2 weeks and was frequently managed with dose reductions. Paraesthesias often had a perioral distribution and were mostly mild in grade, often requiring no therapeutic intervention.<sup>135</sup>

### Mechanisms of resistance to *TRK* inhibitors

As with most TKIs, the emergence of on-target, solventfront or gatekeeper mutations are major mechanisms of acquired resistance to *TRK* inhibitors;<sup>136</sup> nonetheless, off-target activation of downstream pathways was also associated with therapeutic resistance.<sup>137</sup> Next-generation *TRK* inhibitors were developed to address on-target resistance mechanisms such as repotrectinib<sup>138,139</sup> and selitrectinib.<sup>140</sup>

### Suggested approach in *NTRK*-rearranged NSCLC

Whilst drug availability is an issue in some countries, patients with *NTRK*-rearranged metastatic NSCLC should be considered for treatment with a *TRK* inhibitor whenever possible, unless ongoing benefit from standard treatment is evident; entrectinib and larotrectinib are both listed as preferred first-line options for *NTRK*-rearranged NSCLC in the NCCN guidelines.<sup>57</sup> Due to the extremely low frequency of *NTRK* rearrangements in NSCLC, comparisons with standard treatments are difficult; however, treatment with a *TRK* inhibitor should be preferred due to a favourable toxicity profile, higher CNS activity and the achievement of durable responses in most patients.

### Conclusions

*ALK-, ROS1-* and *NTRK*-rearranged tumours represent a distinct clinical and molecular entity amongst NSCLC and have the highest frequency amongst young, non-smoker patients. Identification and proper treatment of *ALK-, ROS1-* and *NTRK-* rearranged tumours with specific inhibitors are critical due to the clinically relevant benefits in QoL, AEs and survival outcomes when compared to standard treatment for fusion-negative, advanced NSCLC.

**Contributions:** AG, DM and MS: conceptualization, writing of original draft and final editing. GM and BR: writing, review and editing. All named authors meet the International Committee of Medical Journal Editors (ICMJE) criteria for authorship for this article, take responsibility for the integrity of the work as a whole and have given their approval for this version to be published. The authors did not receive medical writing assistance.

**Disclosure and potential conflicts of interest:** AJ Gelibter received honoraria for advisory board involvement from Astra Zeneca, MSD, BMS, Roche and Boheringer. The authors declare that they have no other conflicts of interest relevant to this manuscript. The International Committee of Medical Journal Editors (ICMJE) Potential Conflicts of Interests form for the authors is available for download at: https://www.drugsincontext.com/wp-content/uploads/2022/09/dic.2022-3-1-COI.pdf

#### Acknowledgements: None.

Funding declaration: There was no funding associated with the preparation of this article.

**Copyright:** Copyright © 2022 Marinelli D, Siringo M, Metro G, Ricciuti B, Gelibter AJ. Published by *Drugs in Context* under Creative Commons License Deed CC BY NC ND 4.0, which allows anyone to copy, distribute and transmit the article provided it is properly attributed in the manner specified below. No commercial use without permission.

**Correct attribution:** Copyright © 2022 Marinelli D, Siringo M, Metro G, Ricciuti B, Gelibter AJ. https://doi.org/10.7573/dic.2022-3-1. Published by *Drugs in Context* under Creative Commons License Deed CC BY NC ND 4.0.

Article URL: https://www.drugsincontext.com/non-small-cell-lung-cancer-how-to-manage-alk-ros1-and-ntrk-rearranged-disease

**Correspondence:** Alain Gelibter, Division of Medical Oncology B, Policlinico Umberto I, Sapienza – Università di Roma, Rome, Italy. Email: alain.gelibter@uniroma1.it

Provenance: Invited; externally peer reviewed.

Submitted: 3 March 2022; Accepted: 18 August 2022; Publication date: 12 October 2022.

*Drugs in Context* is published by BioExcel Publishing Ltd. Registered office: 6 Green Lane Business Park, 238 Green Lane, New Eltham, London, SE9 3TL, UK.

BioExcel Publishing Limited is registered in England Number 10038393. VAT GB 252 7720 07.

For all manuscript and submissions enquiries, contact the Editorial office editorial@drugsincontext.com

For all permissions, rights and reprints, contact David Hughes david.hughes@bioexcelpublishing.com

### References

- 1. Melosky B, Cheema P, Agulnik J, et al. Canadian perspectives: update on inhibition of ALK-positive tumours in advanced nonsmall-cell lung cancer. *Curr Oncol*. 2018;25(5):317–328. https://doi.org/10.3747/co.25.4379
- Chan BA, Hughes BG. Targeted therapy for non-small cell lung cancer: current standards and the promise of the future. *Transl Lung Cancer Res*. 2015;4(1):36–54. https://doi.org/10.3978/j.issn.2218-6751.2014.05.01
- 3. Wong DW, Leung EL, So KK, et al. The EML4-ALK fusion gene is involved in various histologic types of lung cancers from nonsmokers with wild-type EGFR and KRAS. *Cancer*. 2009;115(8):1723–1733. https://doi.org/10.1002/cncr.24181
- von Laffert M, Warth A, Penzel R, et al. Multicenter immunohistochemical ALK-testing of non–small-cell lung cancer shows high concordance after harmonization of techniques and interpretation criteria. *J Thorac Oncol.* 2014;9(11):1685–1692. https://doi.org/10.1097/JTO.00000000000332
- 5. Rodig SJ, Mino-Kenudson M, Dacic S, et al. Unique clinicopathologic features characterize ALK-rearranged lung adenocarcinoma in the western population. *Clin Cancer Res.* 2009;15(16):5216–5223. https://doi.org/10.1158/1078-0432.CCR-09-0802
- 6. Huang, H. Anaplastic lymphoma kinase (ALK) receptor tyrosine kinase: a catalytic receptor with many faces. *Int J Mol Sci.* 2018;19(11):3448. https://doi.org/10.3390/ijms19113448
- 7. Makimoto G, Ohashi K, Maeda Y, et al. Anaplastic lymphoma kinase fusion: a review of therapeutic drugs and treatment strategies. *Acta Med Okayama*. 2020;74(5):371–379. https://doi.org/10.18926/AMO/60796
- Horn L, Pao W. EML4-ALK: honing in on a new target in non-small-cell lung cancer. J Clin Oncol. 2009;27(26):4232–4235. https://doi.org/10.1200/JCO.2009.23.6661
- 9. Tsao AS, Scagliotti GV, Bunn PA Jr, et al. Scientific advances in lung cancer 2015. *J Thorac Oncol*. 2016;11(5):613–638. https://doi.org/10.1016/j.jtho.2016.03.012
- 10. Guan J, Umapathy G, Yamazaki Y, et al. FAM150A and FAM150B are activating ligands for anaplastic lymphoma kinase. *Elife*. 2015;4:e09811. https://doi.org/10.7554/eLife.09811
- Benayed R, Offin M, Mullaney K, et al. High yield of RNA sequencing for targetable kinase fusions in lung adenocarcinomas with no mitogenic driver alteration detected by DNA sequencing and low tumor mutation burden. *Clin Cancer Res.* 2019;25(15):4712– 4722. https://doi.org/10.1158/1078-0432.CCR-19-0225
- Lazzari C, Spitaleri G, Catania C, et al. Targeting ALK in patients with advanced non small cell lung cancer: biology, diagnostic and therapeutic options. *Crit Rev Oncol Hematol*. 2014;89(3):358–365. https://doi.org/10.1016/j.critrevonc.2013.09.003
- 13. Dacic S, Villaruz LC, Abberbock S, et al. ALK FISH patterns and the detection of ALK fusions by next generation sequencing in lung adenocarcinoma. *Oncotarget*. 2016;7(50):82943–82952. https://doi.org/10.18632/oncotarget.12705
- 14. Friedlaender A, Banna G, Patel S, et al. Diagnosis and treatment of ALK aberrations in metastatic NSCLC. *Curr Treat Options Oncol.* 2019;20(10):79. https://doi.org/10.1007/s11864-019-0675-9
- 15. McCoach CE, Le AT, Gowan K, et al. Resistance mechanisms to targeted therapies in ROS1+ and ALK+ non-small cell lung cancer. *Clin Cancer Res.* 2018;24(14):3334–3347. https://doi.org/10.1158/1078-0432.CCR-17-2452
- McCoach CE, Blakely CM, Banks KC, et al. Clinical utility of cell-free DNA for the detection of ALK fusions and genomic mechanisms of ALK inhibitor resistance in non-small cell lung cancer. *Clin Cancer Res.* 2018;24(12):2758–2770. https://doi.org/10.1158/1078-0432.CCR-17-2588
- 17. Johung KL, Yeh N, Desai NB, et al. Extended survival and prognostic factors for patients with ALK-rearranged non–small-cell lung cancer and brain metastasis. *J Clin Oncol*. 2016;34(2):123–129. https://doi.org/10.1200/JCO.2015.62.0138

- 18. Schrank Z, Chhabra G, Lin L, et al. Current molecular-targeted therapies in NSCLC and their mechanism of resistance. *Cancers*. 2018;10(7):224. https://doi.org/10.3390/cancers10070224
- 19. Kwak EL, Bang YJ, Camidge DR, et al. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med*. 2010;363(18):1693–1703. https://doi.org/10.1056/NEJMoa1006448
- 20. Blackhall F, Ross Camidge D, Shaw AT t al. Final results of the large-scale multinational trial PROFILE 1005: efficacy and safety of crizotinib in previously treated patients with advanced/metastatic ALK-positive non-small-cell lung cancer. *ESMO Open*. 2017;2(3):e000219. https://doi.org/10.1136/esmoopen-2017-000219
- 21. Shaw AT, Kim DW, Nakagawa K, et al. Crizotinib versus chemotherapy in advanced ALK-positive lung cancer. *N Engl J Med*. 2013;368(25):2385–2394. https://doi.org/10.1056/NEJMoa1214886
- 22. Solomon BJ, Mok T, Kim DW, et al. First-line crizotinib versus chemotherapy in ALK-positive lung cancer. *N Engl J Med*. 2014;371(23):2167–2177. https://doi.org/10.1056/NEJMoa1408440
- 23. Costa DB, Kobayashi S, Pandya SS, et al. CSF concentration of the anaplastic lymphoma kinase inhibitor crizotinib. *J Clin Oncol.* 2011;29(15):e443–e445. https://doi.org/10.1200/JCO.2010.34.1313
- 24. Shaw AT, Kim TM, Crinò L, et al. Ceritinib versus chemotherapy in patients with ALK-rearranged non-small-cell lung cancer previously given chemotherapy and crizotinib (ASCEND-5): a randomised, controlled, open-label, phase 3 trial. *Lancet Oncol*. 2017;18(7):874–886. https://doi.org/10.1016/S1470-2045(17)30339-X
- 25. Shaw AT, Kim DW, Mehra R, et al. Ceritinib in ALK-rearranged non-small-cell lung cancer. *N Engl J Med*. 2014;370(13):1189–1197. https://doi.org/10.1056/NEJMoa1311107
- 26. Friboulet L, Li N, Katayama R, Lee CC, et al. The ALK inhibitor ceritinib overcomes crizotinib resistance in non-small cell lung cancer. *Cancer Discov*. 2014;4(6):662–673. https://doi.org/10.1158/2159-8290.CD-13-0846
- 27. Soria JC, Tan DSW, Chiari R, et al. First-line ceritinib versus platinum-based chemotherapy in advanced ALK-rearranged nonsmall-cell lung cancer (ASCEND-4): a randomised, open-label, phase 3 study. *Lancet*. 2017;389(10072):917–929. https://doi.org/10.1016/S0140-6736(17)30123-X
- Claxton L, O'Connor J, Woolacott N, et al. Ceritinib for untreated anaplastic lymphoma kinase-positive advanced non-small-cell lung cancer: an evidence review group evaluation of a NICE single technology appraisal. *Pharmacoeconomics*. 2019;37(5): 645–654. https://doi.org/10.1007/s40273-018-0720-8
- 29. Gadgeel SM, Gandhi L, Riely GJ, et al. Safety and activity of alectinib against systemic disease and brain metastases in patients with crizotinib-resistant ALK-rearranged non-small-cell lung cancer (AF-002JG): results from the dose-finding portion of a phase 1/2 study. *Lancet Oncol*. 2014;15(10):1119–1128. https://doi.org/10.1016/S1470-2045(14)70362-6
- 30. Hida T, Nokihara H, Kondo M, et al. Alectinib versus crizotinib in patients with ALK-positive non-small-cell lung cancer (J-ALEX): an open-label, randomised phase 3 trial. *Lancet*. 2017;390(10089):29–39. https://doi.org/10.1016/S0140-6736(17)30565-2
- 31. Zhou C, Kim SW, Reungwetwattana T, et al. Alectinib versus crizotinib in untreated Asian patients with anaplastic lymphoma kinase-positive non-small-cell lung cancer (ALESIA): a randomised phase 3 study. *Lancet Respir Med.* 2019;7(5):437–446. https://doi.org/10.1016/S2213-2600(19)30053-0
- 32. Peters S, Camidge DR, Shaw AT, et al. Alectinib versus crizotinib in untreated ALK-positive non–small-cell lung cancer. *N Engl J Med.* 2017;377(9):829–838. https://doi.org/10.1056/NEJMoa1704795
- 33. Mok T, Camidge DR, Gadgeel SM, et al. Updated overall survival and final progression-free survival data for patients with treatment-naive advanced ALK-positive non-small-cell lung cancer in the ALEX study. *Ann Oncol.* 2020;31(8):1056–1064. https://doi.org/10.1016/j.annonc.2020.04.478
- 34. Gadgeel S, Peters S, Mok T, et al. Alectinib versus crizotinib in treatment-naive anaplastic lymphoma kinase-positive (ALK+) non-small-cell lung cancer: CNS efficacy results from the ALEX study. *Ann Oncol.* 2018;29(11):2214–2222. https://doi.org/10.1093/annonc/mdy405
- 35. Pérol M, Pavlakis N, Levchenko E, et al. Patient-reported outcomes from the randomized phase III ALEX study of alectinib versus crizotinib in patients with ALK-positive non-small-cell lung cancer. *Lung Cancer*. 2019;138:79–87. https://doi.org/10.1016/j.lungcan.2019.10.002
- 36. Squillace RM, Anjum R, Miller D, et al. AP26113 possesses pan-inhibitory activity versus crizotinib-resistant ALK mutants and oncogenic ROS1 fusions. *Cancer Res.* 2013;73:5655. https://doi.org/10.1158/1538-7445.AM2013-5655
- 37. Kim DW, Tiseo M, Ahn MJ, et al. Brigatinib in patients with Crizotinib-refractory anaplastic lymphoma kinase-positive nonsmall-cell lung cancer: a randomized, multicenter phase II *Trial J Clin Oncol*. 2017;35(22):2490–2498. https://doi.org/10.1200/JCO.2016.71.5904
- Gettinger SN, Bazhenova LA, Langer CJ, et al. Activity and safety of brigatinib in ALK-rearranged non-small-cell lung cancer and other malignancies: a single-arm, open-label, phase 1/2 trial. *Lancet Oncol.* 2016;17(12):1683–1696. https://doi.org/10.1016/S1470-2045(16)30392-8
- 39. Camidge DR, Kim HR, Ahn MJ, et al. Brigatinib versus Crizotinib in ALK-positive non-small-cell lung cancer. *N Engl J Med*. 2018;379(21):2027–2039. https://doi.org/10.1056/NEJMoa1810171

- 40. Camidge DR, Kim HR, Ahn MJ, et al. Brigatinib Versus Crizotinib in ALK inhibitor-naive advanced ALK-positive NSCLC: final results of phase 3 ALTA-1L trial. *J Thorac Oncol.* 2021;16(12):2091–2108. https://doi.org/10.1016/j.jtho.2021.07.035
- 41. Garcia Campelo MR, Lin HM, Zhu Y, et al. Health-related quality of life in the randomized phase III trial of Brigatinib vs Crizotinib in advanced ALK inhibitor-naive ALK+ non-small cell lung cancer (ALTA-1L). *Lung Cancer*. 2021;155:68–77. https://doi.org/10.1016/j.lungcan.2021.03.005
- 42. Horn L, Infante JR, Reckamp KL, et al. Ensartinib (X-396) in ALK-positive non-small cell lung cancer: results from a first-in-human phase I/II, multicenter study. *Clin Cancer Res.* 2018;24(12):2771–2779. https://doi.org/10.1158/1078-0432.CCR-17-2398
- 43. Yang Y, Zhou J, Zhou J, et al. Efficacy, safety, and biomarker analysis of ensartinib in crizotinib-resistant, ALK-positive non-small-cell lung cancer: a multicentre, phase 2 trial. *Lancet Respir Med*. 2020;8(1):45–53. https://doi.org/10.1016/S2213-2600(19)30252-8
- 44. Horn L, Wang Z, Wu G, et al. Ensartinib vs Crizotinib for patients with anaplastic lymphoma kinase-positive non-small cell lung cancer: a randomized clinical trial. *JAMA Oncol*. 2021;7(11):1617–1625. https://doi.org/10.1001/jamaoncol.2021.3523
- 45. Solomon BJ, Besse B, Bauer TM, et al. Lorlatinib in patients with ALK-positive non-small-cell lung cancer: results from a global phase 2 study. *Lancet Oncol.* 2018;19(12):1654–1667. https://doi.org/10.1016/S1470-2045(18)30649-1
- 46. Shaw AT, Bauer TM, de Marinis F, et al. First-line Lorlatinib or Crizotinib in advanced *ALK*-positive lung cancer. *N Engl J Med*. 2020;383(21):2018–2029. https://doi.org/10.1056/NEJMoa2027187
- 47. Bauer TM, Shaw AT, Johnson MLI, et al. Brain penetration of lorlatinib: cumulative incidences of CNS and non-CNS progression with lorlatinib in patients with previously treated ALK-positive non-small-cell lung cancer. *Target Oncol.* 2020;15(1):55–65. https://doi.org/10.1007/s11523-020-00702-4
- 48. Gainor JF, Dardaei L, Yoda S, et al. Molecular mechanisms of resistance to first- and second-generation ALK inhibitors in ALK-rearranged lung cancer. *Cancer Discov*. 2016;6(10):1118–1133. https://doi.org/10.1158/2159-8290.CD-16-0596
- 49. Shaw AT, Friboulet L, Leshchiner I, et al. Resensitization to Crizotinib by the Lorlatinib ALK resistance mutation L1198F. *N Engl J Med.* 2016;374(1):54–61. https://doi.org/10.1056/NEJMoa1508887
- 50. Shaw AT, Solomon BJ, Besse B, et al. ALK resistance mutations and efficacy of Lorlatinib in advanced anaplastic lymphoma kinase-positive non-small-cell lung cancer. *J Clin Oncol*. 2019;37(16):1370–1379. https://doi.org/10.1200/JCO.18.02236
- 51. Yoda S, Lin JJ, Lawrence MS, et al. Sequential ALK inhibitors can select for Lorlatinib-resistant compound *ALK* mutations in ALK-positive lung cancer. *Cancer Discov*. 2018;8(6):714–729. https://doi.org/10.1158/2159-8290.CD-17-1256
- 52. Ou SI, Nagasaka M, Brazel D, et al. Will the clinical development of 4th-generation "double mutant active" ALK TKIs (TPX-0131 and NVL-655) change the future treatment paradigm of ALK+ NSCLC? *Transl Oncol*. 2021;14(11):101191. https://doi.org/10.1016/j.tranon.2021.101191
- 53. Lin JJ, Zhu VW, Yoda S, et al. Impact of EML4-ALK variant on resistance mechanisms and clinical outcomes in ALK-positive lung cancer. *J Clin Oncol*. 2018;36(12):1199–1206. https://doi.org/10.1200/JCO.2017.76.2294
- 54. Yoshida T, Oya Y, Tanaka K, et al. Differential Crizotinib response duration among ALK fusion variants in ALK-positive non-small-cell lung cancer. *J Clin Oncol*. 2016;34(28):3383–3389. https://doi.org/10.1200/JCO.2015.65.8732
- 55. Woo CG, Seo S, Kim SW, et al. Differential protein stability and clinical responses of EML4-ALK fusion variants to various ALK inhibitors in advanced ALK-rearranged non-small cell lung cancer. *Ann Oncol.* 2017;28(4):791–797. https://doi.org/10.1093/annonc/mdw693
- 56. Recondo G, Mezquita L, Facchinetti F, et al. Diverse resistance mechanisms to the third-generation ALK inhibitor lorlatinib in ALK-rearranged lung cancer. *Clin Cancer Res.* 2020;26(1):242–255. https://doi.org/10.1158/1078-0432.CCR-19-1104
- 57. National Comprehensive Cancer Network. Non-small cell lung cancer (Version 3.2022). https://www.nccn.org/professionals/physician\_gls/pdf/nscl.pdf. Accessed July 1, 2022.
- 58. Ota K, Azuma K, Kawahara A, et al. Induction of PD-L1 expression by the EML4-ALK Oncoprotein and downstream signaling pathways in non-small cell lung cancer. *Clin Cancer Res.* 2015;21(17):4014–4021. https://doi.org/10.1158/1078-0432.CCR-15-0016
- 59. Shaw AT, Varghese AM, Solomon BJ, et al. Pemetrexed-based chemotherapy in patients with advanced, ALK-positive non-small cell lung cancer. *Ann Oncol.* 2013;24(1):59–66. https://doi.org/10.1093/annonc/mds242
- 60. Lin JJ, Schoenfeld AJ, Zhu VW, et al. Efficacy of platinum/pemetrexed combination chemotherapy in ALK-positive NSCLC refractory to second-generation ALK inhibitors. *J Thorac Oncol*. 2020;15(2):258–265. https://doi.org/10.1016/j.jtho.2019.10.014
- 61. Socinski MA, Nishio M, Jotte RM, et al. IMpower150 final overall survival analyses for atezolizumab plus bevacizumab and chemotherapy in first-line metastatic nonsquamous NSCLC. *J Thorac Oncol*. 2021;16(11):1909–1924. https://doi.org/10.1016/j.jtho.2021.07.009
- 62. Rikova K, Guo A, Zeng Q, et al. Global survey of phosphotyrosine signaling identifies oncogenic kinases in lung cancer. *Cell*. 2007;131(6):1190–1203. https://doi.org/10.1016/j.cell.2007.11.025
- 63. Bergethon K, Shaw AT, Ou SH, et al. ROS1 rearrangements define a unique molecular class of lung cancers. *J Clin Oncol.* 2012;30(8):863–870. https://doi.org/10.1200/JCO.2011.35.6345

- 64. Parikh DA, Walia G, Freeman-Daily J, et al. Characteristics of patients with ROS1+ cancers: results from the first patientdesigned, global, pan-cancer ROS1 data repository. *JCO Oncol Pract*. 2020;16(2):e183–e189. https://doi.org/10.1200/JOP.19.00135
- 65. Zhang T, Joubert P, Ansari-Pour N, et al. Genomic and evolutionary classification of lung cancer in never smokers. *Nat Genet*. 2021;53(9):1348–1359. https://doi.org/10.1038/s41588-021-00920-0
- 66. Patil T, Smith D, Bunn PA, et al. The incidence of brain metastases in stage IV ROS1-rearranged non-small cell lung cancer and rate of central nervous system progression on crizotinib. *J Thorac Oncol*. 2018;13(11):1717–1726. https://doi.org/10.1016/j.jtho.2018.07.001
- 67. Li Z, Shen L, Ding D, et al. Efficacy of Crizotinib among different types of ROS1 fusion partners in patients with ROS1-rearranged non-small cell lung cancer. *J Thorac Oncol.* 2018;13(7):987–995. https://doi.org/10.1016/j.jtho.2018.04.016
- 68. Chiari R, Ricciuti B, Landi L, et al. ROS1-rearranged non-small-cell lung cancer is associated with a high rate of venous thromboembolism: analysis from a phase II, prospective, multicenter, two-arms trial (METROS). *Clin Lung Cancer*. 2020;21(1): 15–20. https://doi.org/10.1016/j.cllc.2019.06.012
- 69. Alexander M, Pavlakis N, John T, et al. A multicenter study of thromboembolic events among patients diagnosed with ROS1rearranged non-small cell lung cancer. *Lung Cancer*. 2020;142:34–40. https://doi.org/10.1016/j.lungcan.2020.01.017
- 70. Shaw AT, Ou SH, Bang Y, et al. Crizotinib in ROS1-rearranged non-small-cell lung cancer. *N Engl J Med*. 2014;371(21):1963–1971. https://doi.org/10.1056/NEJMoa1406766
- 71. Drilon A, Jenkins C, Iyer S, et al. ROS1-dependent cancers biology, diagnostics and therapeutics. *Nat Rev Clin Oncol.* 2021;18(1):35–55. https://doi.org/10.1038/s41571-020-0408-9
- 72. Lin J, Ritterhouse L, Ali S, et al. ROS1 fusions rarely overlap with other oncogenic drivers in non-small cell lung cancer. *J Thorac Oncol*. 2017;12(5):872–877. https://doi.org/10.1016/j.jtho.2017.01.004
- 73. Mazieres J, A Drilon A, Lusque A, et al. Immune checkpoint inhibitors for patients with advanced lung cancer and oncogenic driver alterations: results from the IMMUNOTARGET registry. *Ann Oncol.* 2019;30(8):1321–1328. https://doi.org/10.1093/annonc/mdz167
- 74. Davies K, Le A, Sheren J, et al. Comparison of molecular testing modalities for detection of ROS1 rearrangements in a Cohort of positive patient samples. *J Thorac Oncol.* 2018;13(10):1474–1482. https://doi.org/10.1016/j.jtho.2018.05.041
- 75. Davies K, Le A, Theodoro M, et al. Identifying and targeting ROS1 gene fusions in non-small cell lung cancer. *Clin Cancer Res.* 2012;18(17):4570–4579. https://doi.org/10.1158/1078-0432.CCR-12-0550
- 76. Sholl L, Sun H, Butaney M, et al. ROS1 immunohistochemistry for detection of ROS1-rearranged lung adenocarcinomas. *Am J Surg Pathol*. 2013;37(9):1441–1449. https://doi.org/10.1097/PAS.0b013e3182960fa7
- 77. Conde E, Hernandez S, Martinez R, et al. Assessment of a new ROS1 immunohistochemistry clone (SP384) for the identification of *ROS1* rearrangements in patients with non–small cell lung carcinoma: the ROSING study. *J Thorac Oncol*. 2019;14(12):2120–2132. https://doi.org/10.1016/j.jtho.2019.07.005
- 78. Cha YJ, Lee JS, Kim HR, et al. Screening of ROS1 rearrangements in lung adenocarcinoma by immunohistochemistry and comparison with ALK rearrangements. *PLoS One*. 2014;9(7):e103333. https://doi.org/10.1371/journal.pone.0103333
- 79. Mescam-Mancini L, Lantuéjoul S, Moro-Sibilot D, et al. On the relevance of a testing algorithm for the detection of ROS1rearranged lung adenocarcinomas. *Lung Cancer*. 2014;83(2):168–173. https://doi.org/10.1016/j.lungcan.2013.11.019
- 80. Yoshida A, Tsuta K, Wakai S, et al. Immunohistochemical detection of ROS1 is useful for identifying ROS1 rearrangements in lung cancers. *Mod Pathol*. 2014;27(5):711–720. https://doi.org/10.1038/modpathol.2013.192
- 81. Boyle TA, Masago K, Ellison KE, et al. ROS1 immunohistochemistry among major genotypes of non-small-cell lung cancer. *Clin Lung Cancer*. 2015;16(2):106–111. https://doi.org/10.1016/j.cllc.2014.10.003
- 82. Shan L, Lian F, Guo L, et al. Detection of ROS1 gene rearrangement in lung adenocarcinoma: comparison of IHC, FISH and realtime RT-PCR. *PLoS One*. 2015;10(3):e0120422. https://doi.org/10.1371/journal.pone.0120422
- 83. Lindeman NI, Cagle PT, Aisner DL, et al. Updated molecular testing guideline for the selection of lung cancer patients for treatment with targeted tyrosine kinase inhibitors: guideline from the college of American pathologists, the international association for the study of lung cancer, and the association for molecular pathology. *Arch Pathol Lab Med*. 2018;142(3):321–346. https://doi.org/10.5858/arpa.2017-0388-CP
- 84. Drilon A, Wang L, Arcila ME, et al. Broad, Hybrid capture-based next-generation sequencing identifies actionable genomic alterations in lung adenocarcinomas otherwise negative for such alterations by other genomic testing approaches. *Clin Cancer Res.* 2015;21(16):3631–3639. https://doi.org/10.1158/1078-0432.CCR-14-2683
- 85. Benayed R, Offin M, Mullaney K, et al. High yield of RNA sequencing for targetable kinase fusions in lung adenocarcinomas with no Mitogenic driver alteration detected by DNA sequencing and low tumor mutation burden. *Clin Cancer Res.* 2019;25(15):4712–4722. https://doi.org/10.1158/1078-0432.CCR-19-0225
- 86. Shaw AT, Riely GJ, Bang YJ, et al. Crizotinib in ROS1-rearranged advanced non-small-cell lung cancer (NSCLC): updated results, including overall survival, from PROFILE 1001. *Ann Oncol.* 2019;30(7):1121–1126. https://doi.org/10.1093/annonc/mdz131

- 87. Wu YL, Yang JC, Kim DW, et al. Phase II study of Crizotinib in East Asian patients with ROS1-positive advanced non-small-cell lung cancer. *J Clin Oncol*. 2018;36(14):1405–1411. https://doi.org/10.1200/JCO.2017.75.5587
- Michels S, Massutí B, Schildhaus HU, et al. Safety and efficacy of Crizotinib in patients with advanced or metastatic ROS1rearranged lung cancer (EUCROSS): a European phase II clinical trial. *J Thorac Oncol*. 2019;14(7):1266–1276. https://doi.org/10.1016/j.jtho.2019.03.020
- 89. Moro-Sibilot D, Cozic N, Pérol M, et al. Crizotinib in c-MET- or ROS1-positive NSCLC: results of the AcSé phase II trial. *Ann Oncol.* 2019;30(12):1985–1991. https://doi.org/10.1093/annonc/mdz407
- 90. Landi L, Chiari R, Tiseo M, et al. Crizotinib in *MET*-deregulated or *ROS1*-rearranged pre-treated non-small cell lung cancer (METROS): a phase II, prospective, multicenter, two-arms trial. *Clin Cancer Res.* 2019;25(24):7312–7319. https://doi.org/10.1158/1078-0432.CCR-19-0994
- 91. Gainor JF, Tseng D, Yoda S, et al. Patterns of metastatic spread and mechanisms of resistance to Crizotinib in *ROS1*-positive nonsmall-cell lung cancer. *JCO Precis Oncol*. 2017;2017:PO.17.00063. https://doi.org/10.1200/PO.17.00063
- 92. Ardini E, Menichincheri M, Banfi P, et al. Entrectinib, a Pan-TRK, ROS1, and ALK inhibitor with activity in multiple molecularly defined cancer indications. *Mol Cancer Ther.* 2016;15(4):628–639. https://doi.org/10.1158/1535-7163.MCT-15-0758
- 93. Menichincheri M, Ardini E, Magnaghi P, et al. Discovery of entrectinib: a new 3-aminoindazole as a potent Anaplastic Lymphoma Kinase (ALK), c-ros oncogene 1 Kinase (ROS1), and Pan-Tropomyosin Receptor Kinases (Pan-TRKs) inhibitor. J Med Chem. 2016;59(7):3392–3408. https://doi.org/10.1021/acs.jmedchem.6b00064
- 94. Drilon A, Siena S, Ou SI, et al. Safety and antitumor activity of the multitargeted Pan-TRK, ROS1, and ALK inhibitor Entrectinib: combined results from two phase I trials (ALKA-372-001 and STARTRK-1). *Cancer Discov*. 2017;7(4):400–409. https://doi.org/10.1158/2159-8290.CD-16-1237
- 95. Drilon A, Siena S, Dziadziuszko R, et al. Entrectinib in ROS1 fusion-positive non-small-cell lung cancer: integrated analysis of three phase 1-2 trials. *Lancet Oncol.* 2020;21(2):261–270. https://doi.org/10.1016/S1470-2045(19)30690-4
- 96. Lim SM, Kim HR, Lee JS, et al. Open-label, multicenter, phase II study of Ceritinib in patients with non-small-cell lung cancer harboring ROS1 rearrangement. *J Clin Oncol*. 2017;35(23):2613–2618. https://doi.org/10.1200/JCO.2016.71.3701
- 97. Zou HY, Li Q, Engstrom LD, et al. PF-06463922 is a potent and selective next-generation ROS1/ALK inhibitor capable of blocking crizotinib-resistant ROS1 mutations. *Proc Natl Acad Sci USA*. 2015;112(11):3493–3498. https://doi.org/10.1073/pnas.1420785112
- 98. Johnson TW, Richardson PF, Bailey S, et al. Discovery of (10R)-7-amino-12-fluoro-2,10,16-trimethyl-15-oxo-10,15,16,17-tetrahydro-2H-8,4-(metheno)pyrazolo[4,3-h][2,5,11]-benzoxadiazacyclotetradecine-3-carbonitrile (PF-06463922), a macrocyclic inhibitor of anaplastic lymphoma kinase (ALK) and c-ros oncogene 1 (ROS1) with preclinical brain exposure and broad-spectrum potency against ALK-resistant mutations. J Med Chem. 2014;57(11):4720–4744. https://doi.org/10.1021/jm500261q
- 99. Shaw AT, Solomon BJ, Chiari R, et al. Lorlatinib in advanced ROS1-positive non-small-cell lung cancer: a multicentre, open-label, single-arm, phase 1-2 trial. *Lancet Oncol.* 2019;20(12):1691–1701. https://doi.org/10.1016/S1470-2045(19)30655-2
- 100. Drilon A, Ou SI, Cho BC, et al. Repotrectinib (TPX-0005) Is a next-generation ROS1/TRK/ALK inhibitor that potently inhibits ROS1/ TRK/ALK Solvent- front mutations. *Cancer Discov*. 2018;8(10):1227–1236. https://doi.org/10.1158/2159-8290.CD-18-0484
- 101. Yun MR, Kim DH, Kim SY, et al. Repotrectinib exhibits potent antitumor activity in treatment-naïve and solvent-front-mutant ROS1rearranged non-small cell lung cancer. *Clin Cancer Res.* 2020;26(13):3287–3295. https://doi.org/10.1158/1078-0432.CCR-19-2777
- 102. Cho BC, Doebele RC, Lin J, et al. MA11.07 phase 1/2 TRIDENT-1 study of repotrectinib in patients with *ROS1*+ or *NTRK*+ advanced solid tumors. *J Thorac Oncol*. 2021;16(3):S174–S175. https://doi.org/10.1016/j.jtho.2021.01.251
- 103. Papadopoulos KP, Borazanci E, Shaw AT, et al. U.S. phase I first-in-human study of Taletrectinib (DS-6051b/AB-106), a ROS1/TRK inhibitor, in patients with advanced solid tumors. *Clin Cancer Res.* 2020;26(18):4785–4794. https://doi.org/10.1158/1078-0432.CCR-20-1630
- 104. Ou SI, Fujiwara Y, Shaw AT, et al. Efficacy of Taletrectinib (AB-106/DS-6051b) in *ROS1*+ NSCLC: an updated pooled analysis of U.S. and Japan phase 1 studies. *JTO Clin Res Rep.* 2020;2(1):100108. https://doi.org/10.1016/j.jtocrr.2020.100108
- 105. Awad MM, Katayama R, McTigue M, et al. Acquired resistance to Crizotinib from a mutation in CD74-ROS1. *N Engl J Med.* 2013;368(25):2395–2401. https://doi.org/10.1056/NEJMoa1215530
- 106. Doebele RC, Dziadziuszko R, Drilon A, et al. LBA28 genomic landscape of entrectinib resistance from ctDNA analysis in STARTRK-2. *Ann Oncol.* 2019;30(Suppl. 5):v865. https://doi.org/10.1093/annonc/mdz394.017
- 107. Lin JJ, Choudhury NJ, Yoda S, et al. Spectrum of mechanisms of resistance to Crizotinib and Lorlatinib in *ROS1* fusion-positive lung cancer. *Clin Cancer Res.* 2021;27(10):2899–2909. https://doi.org/10.1158/1078-0432.CCR-21-0032.
- 108. Song A, Kim TM, Kim DW, et al. Molecular changes associated with acquired resistance to Crizotinib in ROS1-rearranged nonsmall cell lung cancer. *Clin Cancer Res.* 2015;21(10):2379–2387. https://doi.org/10.1158/1078-0432.CCR-14-1350
- 109. McCoach CE, Le AT, Gowan K, et al. Resistance mechanisms to targeted therapies in *ROS1*<sup>+</sup> and *ALK*<sup>+</sup> non-small cell lung cancer. *Clin Cancer Res.* 2018;24(14):3334–3347. https://doi.org/10.1158/1078-0432.CCR-17-2452
- 110. McGranahan N, Swanton C. Clonal heterogeneity and tumor evolution: past, present, and the future. *Cell*. 2017;168(4):613–628. https://doi.org/10.1016/j.cell.2017.01.018

- 111. Kaplan DR, Hempstead BL, Martin-Zanca D, et al. The trk proto-oncogene product: a signal transducing receptor for nerve growth factor. *Science*. 1991;252(5005):554–558. https://doi.org/10.1126/science.1850549
- 112. Klein R, Jing SQ, Nanduri V, et al. The trk proto-oncogene encodes a receptor for nerve growth factor. *Cell*. 1991;65(1):189–197. https://doi.org/10.1016/0092-8674(91)90419-y
- 113. Klein R, Nanduri V, Jing SA, et al. The trkB tyrosine protein kinase is a receptor for brain-derived neurotrophic factor and neurotrophin-3. *Cell*. 1991;66(2):395–403. https://doi.org/10.1016/0092-8674(91)90628-c
- 114. Lamballe F, Klein R, Barbacid M. trkC, a new member of the trk family of tyrosine protein kinases, is a receptor for neurotrophin-3. *Cell*. 1991;66(5):967–979. https://doi.org/10.1016/0092-8674(91)90442-2
- 115. Knezevich SR, McFadden DE, Tao W, et al. A novel ETV6-NTRK3 gene fusion in congenital fibrosarcoma. *Nat Genet*. 1998;18(2):184–187. https://doi.org/10.1038/ng0298-184
- 116. Cocco E, Scaltriti M, Drilon A. NTRK fusion-positive cancers and TRK inhibitor therapy. *Nat Rev Clin Oncol*. 2018;15(12):731–747. https://doi.org/10.1038/s41571-018-0113-0
- 117. Rosen EY, Goldman DA, Hechtman JF, et al. TRK fusions are enriched in cancers with uncommon histologies and the absence of canonical driver mutations. *Clin Cancer Res.* 2020;26(7):1624–1632. https://doi.org/10.1158/1078-0432.CCR-19-3165
- 118. Forsythe A, Zhang W, Phillip Strauss U, et al. A systematic review and meta-analysis of neurotrophic tyrosine receptor kinase gene fusion frequencies in solid tumors. *Ther Adv Med Oncol.* 2020;12:1758835920975613. https://doi.org/10.1177/1758835920975613
- 119. Farago AF, Taylor MS, Doebele RC, et al. Clinicopathologic Features of non–small-cell lung cancer Harboring an *NTRK* gene fusion. *JCO Precis Oncol.* 2018;2018:PO.18.00037. https://doi.org/10.1200/PO.18.00037
- 120. Vaishnavi A, Capelletti M, Le AT, et al. Oncogenic and drug-sensitive NTRK1 rearrangements in lung cancer. *Nat Med*. 2013;19(11):1469–1472. https://doi.org/10.1038/nm.3352
- 121. Martin-Zanca D, Hughes SH, Barbacid M. A human oncogene formed by the fusion of truncated tropomyosin and protein tyrosine kinase sequences. *Nature*. 1986;319(6056):743–748. https://doi.org/10.1038/319743a0
- 122. Reinach FC, MacLeod AR. Tissue-specific expression of the human tropomyosin gene involved in the generation of the trk oncogene. *Nature*. 1986;322(6080):648–650. https://doi.org/10.1038/322648a0
- 123. Coulier F, Martin-Zanca D, Ernst M, et al. Mechanism of activation of the human trk oncogene. *Mol Cell Biol*. 1989;9(1):15–23. https://doi.org/10.1128/mcb.9.1.15-23.1989
- 124. Westphalen CB, Krebs MG, Le Tourneau C, et al. Genomic context of NTRK1/2/3 fusion-positive tumours from a large real-world population. *NPJ Precis Oncol*. 2021;5(1):69. https://doi.org/10.1038/s41698-021-00206-y
- 125. Marchiò C, Scaltriti M, Ladanyi M, et al. ESMO recommendations on the standard methods to detect NTRK fusions in daily practice and clinical research. Ann Oncol. 2019;30(9):1417–1427. https://doi.org/10.1093/annonc/mdz204
- 126. Park HJ, Baek I, Cheang G, et al. Comparison of RNA-based next-generation sequencing assays for the detection of NTRK gene fusions. *J Mol Diagn*. 2021;23(11):1443–1451. https://doi.org/10.1016/j.jmoldx.2021.07.027
- 127. Cohen D, Hondelink LM, Solleveld-Westerink N, et al. Optimizing mutation and fusion detection in NSCLC by sequential DNA and RNA sequencing. *J Thorac Oncol*. 2020;15(6):1000–1014. https://doi.org/10.1016/j.jtho.2020.01.019
- 128. Hechtman JF, Benayed R, Hyman DM, et al. Pan-Trk immunohistochemistry is an efficient and reliable screen for the detection of NTRK fusions. *Am J Surg Pathol*. 2017;41(11):1547–1551. https://doi.org/10.1097/PAS.00000000000911
- 129. Solomon JP, Linkov I, Rosado A, et al. NTRK fusion detection across multiple assays and 33,997 cases: diagnostic implications and pitfalls. *Mod Pathol*. 2020;33(1):38–46. https://doi.org/10.1038/s41379-019-0324-7
- 130. Drilon A, Laetsch TW, Kummar S, et al. Efficacy of larotrectinib in TRK fusion-positive cancers in adults and children. *N Engl J Med*. 2018 22;378(8):731–739. https://doi.org/10.1056/NEJMoa1714448
- 131. Drilon A, Tan DSW, Lassen UN, et al. Efficacy and safety of larotrectinib in patients with tropomyosin receptor kinase fusion-positive lung cancers. JCO Precis Oncol. 2022;6:e2100418. https://doi.org/10.1200/PO.21.00418
- 132. Doebele RC, Drilon A, Paz-Ares L, et al. Entrectinib in patients with advanced or metastatic NTRK fusion-positive solid tumours: integrated analysis of three phase 1-2 trials. *Lancet Oncol*. 2020;21(2):271–282. https://doi.org/10.1016/S1470-2045(19)30691-6
- 133. Drilon A, Paz-Ares L, Doebele RC, et al. Entrectinib in NTRK fusion-positive NSCLC: updated integrated analysis of STARTRK-2, STARTRK-1 and ALKA-372-001. *Ann Oncol.* 2020;31:S474–S475. https://doi.org/10.1016/j.annonc.2020.08.657
- 134. Dziadziuszko R, Siena S, Tan DSW, et al. Efficacy of entrectinib in patients with NTRK or ROS1 fusion-positive NSCLC with CNS metastases at baseline. *Ann Oncol.* 2020;31:S833–S834. https://doi.org/10.1016/j.annonc.2020.08.1602
- 135. Liu D, Flory J, Lin A, et al. Characterization of on-target adverse events caused by TRK inhibitor therapy. *Ann Oncol.* 2020;31(9):1207–1215. https://doi.org/10.1016/j.annonc.2020.05.006
- 136. Drilon A, Nagasubramanian R, Blake JF, et al. A next-generation TRK kinase inhibitor overcomes acquired resistance to prior TRK kinase inhibition in patients with TRK fusion-positive solid tumors. *Cancer Discov.* 2017;7(9):963–972. https://doi.org/10.1158/2159-8290.CD-17-0507

- 137. Cocco E, Schram AM, Kulick A, et al. Resistance to TRK inhibition mediated by convergent MAPK pathway activation. *Nat Med*. 2019;25(9):1422–1427. https://doi.org/10.1038/s41591-019-0542-z
- 138. Drilon A, Ou SI, Cho BC, et al. Repotrectinib (TPX-0005) is a next-generation ROS1/TRK/ALK inhibitor that potently inhibits ROS1/ TRK/ALK solvent- front mutations. *Cancer Discov*. 2018;8(10):1227–1236. https://doi.org/10.1158/2159-8290.CD-18-0484
- 139. Murray BW, Rogers E, Zhai D, et al. Molecular characteristics of repotrectinib that enable potent inhibition of TRK fusion proteins and resistant mutations. *Mol Cancer Ther.* 2021;20(12):2446–2456. https://doi.org/10.1158/1535-7163.MCT-21-0632
- 140. Hemming ML, Nathenson MJ, Lin JR, et al. Response and mechanisms of resistance to larotrectinib and selitrectinib in metastatic undifferentiated sarcoma harboring oncogenic fusion of NTRK1. *JCO Precis Oncol*. 2020;4:79–90. https://doi.org/10.1200/po.19.00287